

**THE EFFECT OF STATISTICAL ERROR MODEL  
FORMULATION ON THE FIT AND SELECTION OF  
MATHEMATICAL MODELS OF TUMOR GROWTH  
FOR SMALL SAMPLE SIZES**

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**Abstract:** When fitting a mathematical model to a given data set using inverse problems, the correctness of both the mathematical model and the statistical error models are important since an incorrect statistical or observational model directly affects both the estimates and their corresponding standard errors. The effects of these models, among many other factors, are dependent on the *sample size* and the *information content of the data set*.

In this article, we investigate how the choice of the statistical error model affects the mathematical model fit and accuracy of parameter estimates in small sample size tumor growth data sets. We specifically seek to determine the appropriate statistical error model for *small sample size* breast, lung and HPV tumor growth *data sets* obtained from studies on mice.

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We find that for small sample sizes the selection of the best statistical error model is not straightforward and requires the examination of multiple criteria for model fit and uncertainty. Therefore, selection of the best mathematical model is not an easy process for small sample size tumor data and selection of a model based on few data points may not prove accurate. We encourage further research on the optimal design of experiments (duration and number of observations) in order to best fit mathematical models to tumor growth data.

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**Key Words:** tumor growth models, mathematical and statistical models, sensitivity, complex-step method, residual analysis

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## 1. Introduction

Some standard and simple mathematical models are commonly used in tumor growth modeling and prediction studies ([9, 13, 14, 19, 23, 21, 25, 26]). A rather striking commonality in most of these studies is the small longitudinal nature (i.e., in terms of the duration and number  $N$  of observations or time points) of the data sets employed for model validation.

Our primary goal is to better understand statistical error models that arise in the observation process for tumor data collection. As explained in [5, 8] and below, in order to verify and select the best mathematical model, the choice of an appropriate statistical error model is critical. In this study, we examine how the choice of statistical error model (and hence the form of least squares employed in the inverse problem) affects the accuracy and uncertainty of the mathematical model fits to data. We investigate this in a selection for tumor growth data from studies on mice with a small number of sampling observations [9, 14]. Thus our effort can be considered a further step in the goals of the authors of [9] in their efforts to more fully understand the form of the observation errors in tumor data collection processes.

In our study, we use the same suite of mathematical models as discussed in [9]. These are the logistic, Gompertz, power law, exponential linear, generalized logistic, dynamic carrying capacity, and Von Bertalanffy models summarized for completeness in Section 2.1 along with the data we have chosen to illustrate our efforts in Section 2.2. Using several sets of tumor growth data from [9] and [14], we estimate model parameters for the mathematical models via the inverse problem methodology summarized in Section 3. This methodology includes the complex-step approach to sensitivity and standard error (SE) computation, residual analysis, second order differencing applied directly to the

data, and Akaike-based information criteria selection procedures. In Section 4 we study the model solutions and the precision of the parameter estimates to investigate how the choice of the statistical error model affects the results of the mathematical models. Moreover, we estimate model parameters using a family of statistical error models and use various computational tools to attempt to determine the ones appropriate for such data sets. Finally in Section 4.4 we use Akaike information criterion (as introduced in Section 3.3) in attempts to rank order our models with the various data sets. We conclude in Section 5 with our conclusions and call for further design-of-experiments investigations.

## 2. Models and Data Sets

### 2.1. Review of Tumor Growth Models

The following are the seven tumor growth mathematical models we will use in our study. These are the same models proposed, carefully explained, and used in [9]. Readers can find a detailed explanation of the rationale for possible use of one or more of these various mathematical models in describing tumor growth in [9]. Here we will be interested in their use in the context of determining an appropriate corresponding statistical error model that can lead to reasonable levels of uncertainty quantification of parameter estimates based on data sets typically available to modelers.

#### 2.1.1. Logistic Model

The Logistic model is given by

$$\begin{cases} \frac{dV}{dt} = aV \left(1 - \frac{V}{K}\right), & t > 0, \\ V(0) = V_0, \end{cases}$$

where  $V$  represents tumor volume,  $a$  is the growth rate, and  $K$  is the carrying capacity. The analytic solution is given by

$$V(t) = \frac{KV_0e^{at}}{K + V_0(e^{at} - 1)}.$$

The parameters of interest in fitting this model to data are  $\boldsymbol{\theta} = (\mathbf{q}, V_0) = (a, K, V_0)$ .

### 2.1.2. Gompertz Model

The Gompertz model is given by

$$\begin{cases} \frac{dV}{dt} = ae^{-\beta t}V, & t > 0 \\ V(0) = V_0, \end{cases}$$

where  $V$  represents tumor volume,  $a$  is the growth rate, and  $\beta$  is the rate of exponential decay of the growth rate. The analytic solution is given by

$$V(t) = V_0 e^{\frac{a}{\beta}(1-e^{-\beta t})}.$$

The parameters of interest in fitting this model to data are  $\theta = (\mathbf{q}, V_0) = (a, \beta, V_0)$ .

### 2.1.3. Generalized Logistic Model

The generalized logistic model is given by

$$\begin{cases} \frac{dV}{dt} = aV \left(1 - \left(\frac{V}{K}\right)^\nu\right), & t > 0, \\ V(0) = V_0, \end{cases}$$

where  $V$  represents volume,  $a$  is the growth rate,  $K$  is the carrying capacity, and  $\nu$  is a constant. When  $\nu = 1$  we have the logistic model, whereas when  $\nu > 1$  the model demonstrates an exponential growth curve and decelerates quickly as  $K$  is maximized [23]. The analytic solution of the system is given by

$$V(t) = \frac{KV_0}{(V_0^\nu + (K^\nu - V_0^\nu)e^{-avt})^{1/\nu}}.$$

The parameters of interest in fitting this model to data are  $\theta = (\mathbf{q}, V_0) = (a, K, \nu, V_0)$ .

### 2.1.4. Dynamic Carrying Capacity Model

The dynamic carrying capacity model is given by

$$\begin{cases} \frac{dV}{dt} = aV \log\left(\frac{K}{V}\right), & t > 0, \\ \frac{dK}{dt} = bV^{2/3}, & t > 0, \\ V(0) = V_0, \quad K(0) = K_0, \end{cases} \quad (1)$$

where  $V$  represents tumor volume,  $a$  is the growth rate,  $K$  is carrying capacity, and  $b$  is the growth or decay rate of the carrying capacity. The parameters of interest in fitting this model to data are  $\theta = (\mathbf{q}, V_0, K_0) = (a, b, V_0, K_0)$ .

### 2.1.5. Power Law Model

The power law model is given by

$$\begin{cases} \frac{dV}{dt} = aV^\mu, \\ V(0) = V_0, \end{cases}$$

where  $V$  represents tumor volume,  $a$  is the growth rate, and  $\mu$  is the allometry volume factor. The analytic solution is given by

$$V(t) = \left( at(1 - \mu) + V_0^{1-\mu} \right)^{\frac{1}{1-\mu}}.$$

The parameters of interest in fitting this model to data are  $\theta = (\mathbf{q}, V_0) = (a, \mu, V_0)$ . Note that the power law model is a special case of the Von Bertalanffy model below, in which we let  $b = 0$ .

### 2.1.6. Von Bertalanffy Model

The Von Bertalanffy model is given by

$$\begin{cases} \frac{dV}{dt} = aV^\mu - bV, \\ V(0) = V_0, \end{cases}$$

where  $V$  represents tumor volume,  $a$  is the growth rate, and  $b$  is the term corresponding to the loss of volume. The analytic solution is given by

$$V(t) = \left( \frac{a}{b} + \left( V_0^{1-\mu} - \frac{a}{b} \right) e^{-b(1-\mu)t} \right)^{\frac{1}{1-\mu}}.$$

The parameters of interest in fitting this model to data are  $\theta = (\mathbf{q}, V_0) = (a, b, \mu, V_0)$ .

### 2.1.7. Exponential Linear Model

The exponential linear model is given by

$$\begin{cases} \frac{dV}{dt} = a_0V, & t \leq \tau, \\ \frac{dV}{dt} = a_1, & t > \tau, \\ V(0) = V_0, \end{cases}$$

where  $V$  represents tumor volume,  $a_0$  is the exponential growth rate, and  $a_1$  is the linear growth rate. The analytic solution is given by

$$V(t) = \begin{cases} V_0 e^{a_0 t} & t \leq \tau, \\ a_1(t - \tau) + V_0 e^{a_0 \tau} & t > \tau, \end{cases}$$

where  $\tau = \frac{1}{a_0} \ln\left(\frac{a_1}{a_0 V_0}\right)$  when  $V(t)$  is continuously differentiable. The parameters of interest in fitting this model to data are  $\boldsymbol{\theta} = (\mathbf{q}, V_0) = (a_0, a_1, V_0)$ .

## 2.2. Data Sets

Data from three different types of tumors were utilized in this work: breast, lung, and skin (HPV) tumors. The lung and breast tumor data were obtained from [9] and the HPV data came from [14]. All the data were collected in labs on mice and extracted from figures in [9, 14] using the data extraction tools Data Thief (Version III) and WebPlotDigitizer (Version 3.12) [20, 24].

### 2.2.1. Data Set Descriptions

For the lung tumor data, male mice were injected subcutaneously with Murine Lewis lung carcinoma after the cells were cultured in a high glucose DMEM. For the breast tumor data, human breast carcinoma  $LM2 - 4^{LUC+}$  was implanted into the mammary fat pads of female immunosuppressed mice. The day the cancer cells were injected/implanted was considered day 0 and subsequent measurements of the tumors were collected using calipers to subcutaneously measure the largest ( $L$ ) and smallest ( $w$ ) diameters of the tumor. Assuming an ellipsoid shape, the volume of the tumor was then calculated using the formula

$$V = \frac{\pi}{6} w^6 L.$$

Lung tumors were allowed to grow to a maximum volume of  $1.5\text{cm}^3$  before the mouse was euthanized, resulting in data collected over a 4 to 22 day period with volumes ranging from  $14 - 1492\text{mm}^3$ . Two experiments were conducted on ten mice each, resulting in twenty lung tumor data sets. Breast tumors were allowed to grow to a maximum volume of  $2\text{cm}^3$  before the mouse was euthanized, resulting in data collected over an 18 to 38 day period with volumes ranging from  $202 - 1902\text{mm}^3$ . Experiments were conducted on a total of 34 mice tumors, resulting in 34 breast tumor data sets [9].

For the skin (HPV) tumors, mice were bred to carry E6/E7 double transgenes and treated with DMBA/TPA. Experiments were conducted on 8 mice.

For each mouse, three tumors were randomly selected and measurements were taken biweekly resulting in twenty-four data sets for the skin (HPV) tumors. Measurements began when the tumor became visible (approximately 1 – 2mm in diameter) with day 0 set to the date of the first measurement. Each tumor's length ( $x$ ) and width ( $y$ ) were measured three times using measuring calipers and the medians ( $M_x$  and  $M_y$ ) of these measurements were calculated. Assuming an ellipsoid shape, the volume of the tumor was calculated using the formula

$$V = \frac{\pi}{6}(M_x M_y)^{3/2}.$$

Skin tumors were allowed to grow to a maximum diameter of 1cm before the mouse was euthanized. Life spans of individual mice ranged from 42 to 105 days. Tumor size, at its largest, ranges from 295.55mm<sup>3</sup> to 864.82mm<sup>3</sup> [14].

In this study, we examined all combinations of the proposed mathematical models and a selection of representative statistical models on a subset of tumors from each data set. We selected four mice of each tumor type to analyze the model fits and choice of statistical error model. We chose the following tumors to show how different models fit the data sets with varying dynamics:

1. *Breast Tumors 15, 16, 31, and 33.* Breast Tumor 15 (B15) has 8 data points and time ranges from day 18 to day 34. Breast Tumor 16 (B16) has 8 data points and time ranges from day 18 to day 34. Breast Tumor 31 (B31) has 7 data points and time ranges from day 22 to day 36. Breast Tumor 33 (B33) has 8 data points and time ranges from day 22 to day 38.
2. *Lung Tumors 2, 4, 5, and 9.* Lung Tumor 2 (L2) has 11 data points and time ranges from day 5 to day 20. Lung Tumor 4 (L4) has 12 data points and time ranges from day 6 to day 22. Lung Tumor 5 (L5) has 11 data points and time ranges from day 5 to day 20. Lung Tumor 9 (L9) has 10 data points and time ranges from day 6 to day 20.
3. *Skin (HPV) Tumors 4, 9, 12, and 24.* Tumor 4 (T4) has 23 data points and time ranges from day 0 to day 76. Tumor 9 (T9) has 24 data points and time ranges from day 0 to day 88. Tumor 12 (T12) has 14 data points and time ranges from day 0 to day 52. Tumor 24 (T24) has 13 data points and time ranges from day 0 to day 42.

### 3. Components of Inverse Problem Studies: Mathematical and Statistical Models

Given a set of observations, the process of estimating model parameters is termed an inverse problem. Following the expositions in [5, 7, 8], we present a short description of the methodology of the inverse problem for the following general 1-dimensional dynamical model:

$$\begin{aligned}\frac{dx}{dt}(t) &= g(t, x(t); \mathbf{q}), \\ x(t_0) &= x_0,\end{aligned}$$

with observation process

$$f(t; \boldsymbol{\theta}) = \mathcal{C}x(t; \boldsymbol{\theta}),$$

where  $\boldsymbol{\theta} = [\mathbf{q}^T, x_0^T]^T \in \mathbb{R}^{k_\theta}$ ,  $\mathbf{q} \in \mathbb{R}^{k_q}$  is a vector parameter and  $\mathcal{C}$  is the observation constant.

Suppose we have  $N$  longitudinal observations  $\{Y_j\}$  (with realizations  $\{y_j\}$ ) corresponding to  $f(t_j; \boldsymbol{\theta}) = \mathcal{C}x(t_j; \boldsymbol{\theta})$ ,  $j = 1, \dots, N$ , and let  $\Omega_\theta$  be the set of admissible parameters. In general,  $\{Y_j\}$  will not be exactly equal to  $f(t_j, \boldsymbol{\theta})$ . To account for this uncertainty in the observations, we use a statistical error model for the observation process given by

$$Y_j = f(t_j; \boldsymbol{\theta}_0) + f^\gamma(t_j; \boldsymbol{\theta}_0)\mathcal{E}_j, \quad j = 1, \dots, N, \quad \gamma \geq 0. \quad (2)$$

Here,  $\boldsymbol{\theta}_0 \in \Omega_\theta$  represents the “true” or nominal parameter set that generates  $\{Y_j\}$  (assumed to exist), and  $f^\gamma(t_j; \boldsymbol{\theta}_0)\mathcal{E}_j$  represents the measurement error or some other phenomena that causes  $Y_j \neq f(t_j; \boldsymbol{\theta}_0)$ . We assume  $\mathcal{E}_j$  are independent over time with mean zero and constant variance.

When  $\gamma = 0$ , i.e., when the error has constant variance, the statistical error model (2) is called the *absolute error model* and we estimate the parameters by minimizing the ordinary least squares (OLS) cost function:

$$\boldsymbol{\theta}_{\text{OLS}} = \arg \min_{\boldsymbol{\theta} \in \Omega_\theta} \sum_{j=1}^N (Y_j - f(t_j; \boldsymbol{\theta}))^2,$$

with realization

$$\hat{\boldsymbol{\theta}}_{\text{OLS}} = \arg \min_{\boldsymbol{\theta} \in \Omega_\theta} \sum_{j=1}^N (y_j - f(t_j; \boldsymbol{\theta}))^2. \quad (3)$$

When  $\gamma \neq 0$ , i.e., when the error has non-constant variance, we use an iterated reweighted weighted least squares (IRWLS) scheme to estimate the

parameters. Note, when  $\gamma = 1$ , the statistical error model is called the *relative error model*. We compute the parameter estimates by solving

$$\boldsymbol{\theta}_{\text{IRWLS}} = \arg \widetilde{\min}_{\boldsymbol{\theta} \in \Omega_{\boldsymbol{\theta}}} \sum_{j=1}^N \frac{1}{f^{2\gamma}(t_j; \boldsymbol{\theta})} (Y_j - f(t_j; \boldsymbol{\theta}))^2,$$

where  $\widetilde{\min}$  denotes an iterative minimization scheme that is explained in Section 3.1.1. Note, this scheme is not equivalent to a traditional minimization. To obtain realizations of  $\boldsymbol{\theta}_{\text{IRWLS}}$ , we solve

$$\hat{\boldsymbol{\theta}}_{\text{IRWLS}} = \arg \widetilde{\min}_{\boldsymbol{\theta} \in \Omega_{\boldsymbol{\theta}}} \sum_{j=1}^N \frac{1}{f^{2\gamma}(t_j; \boldsymbol{\theta})} (y_j - f(t_j; \boldsymbol{\theta}))^2. \quad (4)$$

For further and more in depth discussion on inverse problem methodology, we refer the reader to [5, 7, 8, 11, 12, 22].

### 3.1. Computational Methods

To fit the seven mathematical models to the data, we solve an inverse problem in each case to estimate model parameters. When an absolute statistical error model is used, we estimate  $\hat{\boldsymbol{\theta}}_{\text{OLS}}$  using Matlab's built-in optimization solver *fmincon* directly to minimize the cost function in (3). In the case when  $\gamma > 0$ , we use the numerical algorithm provided in [5, 7] to estimate  $\hat{\boldsymbol{\theta}}_{\text{IRWLS}}$  which is summarized below.

#### 3.1.1. IRWLS Algorithm

To compute  $\hat{\boldsymbol{\theta}}_{\text{IRWLS}}$ , we will make use of the weighted least squares cost functional:

$$\hat{\boldsymbol{\theta}} = \arg \min_{\boldsymbol{\theta} \in \Omega_{\boldsymbol{\theta}}} \sum_{j=1}^N w_j (y_j - f(t_j; \boldsymbol{\theta})) \quad (5)$$

1. Solve for an initial estimate  $\hat{\boldsymbol{\theta}}^0$  by solving (3). Set  $l = 0$ .
2. Compute the weights  $w_j = f^{-2\gamma}(t_j; \hat{\boldsymbol{\theta}}^{(l)})$ .
3. Re-estimate  $\hat{\boldsymbol{\theta}}$  by solving (5) with the weights computed in step 2 fixed to obtain  $\hat{\boldsymbol{\theta}}^{(l+1)}$ .

4. If  $\hat{\theta}^{(l)}$  and  $\hat{\theta}^{(l+1)}$  are sufficiently close, set  $\hat{\theta}_{\text{IRWLS}} = \hat{\theta}^{(l+1)}$ . Otherwise, set  $l = l + 1$  and return to step 2.

All of the dynamical systems are solved using the Matlab `ode45` ODE solver which is based on the fourth order Runge-Kutta scheme.

### 3.1.2. The Complex-Step Method for Computing Sensitivities

There are numerous methods for computing sensitivities with respect to parameters in models such as those under consideration here. These are discussed in some detail in [5, 8] and include forward differences, sensitivity equations, traditional sensitivity functions (TSF), relative sensitivity functions (RSF), and automatic differentiation (see [5, p. 74-76]). Here we use a method originally based on analyticity [15, 16, 17, 18] of the functions involved. The method was recently developed further in [1, 2].

This method, referred to as *the complex-step method*, as derived earlier relied on the Cauchy-Riemann equations in a crucial way. More recently in [1, 2] the method was shown to be much more widely applicable. Indeed one can argue for any  $C^2$  function in the parameter  $\theta$ , one has the 2nd order Taylor expansion using the complex step  $ih$

$$f(t; \theta + ih) = f(t; \theta) + ih \frac{\partial f}{\partial \theta}(t; \theta) + \mathcal{R}_2(t; \theta; h) \quad (6)$$

where

$$\lim_{h \rightarrow 0} \frac{\mathcal{R}_2(h)}{h} = 0.$$

Taking the imaginary parts of both sides and dividing by  $h$  in (6) results in the first order approximation

$$\frac{\partial f}{\partial \theta}(t; \theta) = f'(t; \theta) \approx \frac{\text{Im}[f(t; \theta + ih)]}{h} \quad (7)$$

with a truncation error  $E_T$  approximated by

$$E_T(h) \approx \frac{h^2}{6} \frac{\partial^3 f}{\partial \theta^3}(t; \theta).$$

Terms of order  $h^2$  and higher can be ignored because the step size  $h$  can be chosen up to machine precision. The method is accurate down to a specific step size we can call  $h_{\text{crit}}$ . Below  $h_{\text{crit}}$ , underflow occurs and the approximation becomes useless. This derivative estimate constitutes a big advantage

over other approximation approaches to sensitivities. First, it is applicable for problems with less smoothness than analyticity (e.g., functions only  $C^2$  in the parameters). Moreover, usual finite-difference approximations are subject to *subtractive error* due to the differencing operation. On the other hand, the accuracy of the complex-step estimates is *only limited by the numerical precision of the algorithm that evaluates the function  $f$* .

### 3.1.3. The Complex-Step Method: Implementation

General steps for implementing the complex-step method for computing  $df/dx$  are

1. Define all functions and operators that are not defined for complex arguments. For example *max*, *min* and *abs*.
2. Add a small complex step  $ih$  to the desired variable  $x$ , run the algorithm that evaluates  $f$ .
3. Compute  $df/dx$  using  $\frac{df}{dx} \approx \frac{\text{Im}[f(x+ih)]}{h}$

### 3.1.4. Computation of Standard Errors

**Absolute error:** We compute the sensitivity matrix

$$\chi_{j,k} = \frac{\partial f(t_j, \hat{\boldsymbol{\theta}})}{\partial \hat{\boldsymbol{\theta}}_k}, \quad j = 1, \dots, N, \quad k = 1, \dots, \kappa_{\boldsymbol{\theta}}, \quad (8)$$

which is done using the complex-step method as detailed above and in [1]. Note that  $\chi = \chi^N$  is an  $N \times \kappa_{\boldsymbol{\theta}}$  matrix. The constant variance is estimated by

$$\hat{\sigma}^2 = \frac{1}{N - \kappa_{\boldsymbol{\theta}}} \left[ \sum_{j=1}^N [y_j - f(t_j, \hat{\boldsymbol{\theta}})]^2 \right].$$

The covariance matrix is approximately given by

$$\Sigma_0^N \approx \sigma_0^2 [\chi^T(\boldsymbol{\theta}_0)\chi(\boldsymbol{\theta}_0)]^{-1},$$

and the approximate Fisher Information Matrix (FIM) is given by

$$F = (\Sigma_0^N)^{-1}. \quad (9)$$

As  $\theta_0$  and  $\sigma_0^2$  are unknown, the covariance matrix is estimated by

$$\hat{\Sigma}^N(\hat{\theta}) = \hat{\sigma}^2 [\chi^T(\hat{\theta})\chi(\hat{\theta})]^{-1}, \quad (10)$$

for which the corresponding estimate of the FIM is

$$\hat{F} = (\hat{\Sigma}^N(\hat{\theta}))^{-1} = \frac{1}{\hat{\sigma}^2} [\chi^T(\hat{\theta})\chi(\hat{\theta})]. \quad (11)$$

Then, the asymptotic standard errors are estimated by

$$SE_k(\hat{\theta}) = \sqrt{(\hat{\Sigma}^N(\hat{\theta}))_{kk}}, \quad k = 1, \dots, \kappa_{\theta}. \quad (12)$$

**Relative error:** For the generalized weighted least squares formulations (IR-WLS) used here, we may define the standard errors by the formula

$$SE_k = \sqrt{\hat{\Sigma}_{kk}^N(\hat{\theta})}, \quad k = 1, \dots, \kappa_{\theta},$$

where the covariance matrix  $\hat{\Sigma}^N$  is given by

$$\hat{\Sigma}^N(\hat{\theta}) = \hat{\sigma}^2 (\chi^T(\hat{\theta})W(\hat{\theta})\chi(\hat{\theta}))^{-1} = \hat{\sigma}^2 \hat{F}^{-1}.$$

Here

$$\hat{F} = [\chi^T(\hat{\theta})W(\hat{\theta})\chi(\hat{\theta})] \quad (13)$$

is the Fisher Information Matrix defined in terms of the sensitivity matrix defined above and written compactly as

$$\chi = \frac{\partial f}{\partial \theta} = \left( \frac{\partial f(t_1; \hat{\theta})}{\partial \theta}, \dots, \frac{\partial f(t_N; \hat{\theta})}{\partial \theta} \right)$$

of size  $N \times \kappa_{\theta}$  (recall  $N$  is the number of data points and  $\kappa_{\theta}$  is the number of estimated parameters) and the weight matrix  $W$  is defined by

$$W^{-1}(\hat{\theta}) = \text{diag}(f(t_1; \hat{\theta})^{2\gamma}, \dots, f(t_N; \hat{\theta})^{2\gamma}).$$

We use the approximation of the variance given by

$$\sigma_0^2 \approx \hat{\sigma}(\hat{\theta})^2 = \frac{1}{N - \kappa_{\theta}} \sum_{i=1}^N \frac{1}{f(t_i; \hat{\theta})^{2\gamma}} (y_i - f(t_i; \hat{\theta}))^2.$$

A quick comparison of (11) and (13) reveals that the uncertainty calculations *will be invalid if one chooses an incorrect statistical error model* to compute the corresponding standard errors. This of course also affects directly the calculations of the estimates themselves.

### 3.2. Methods for Selection of a Statistical Error Model

Before estimating model parameters, one should make efforts to determine the correct statistical error model to account for the uncertainty of the data (such as observation error). We note that a misspecified statistical error model can lead to an incorrect estimation of the parameters as well as their uncertainty bounds. With the assumption on the errors  $\mathcal{E}_j$  (i.e., *i.i.d.*, mean zero, and constant variance) in the previous section, there are generally two ways of selecting the correct statistical error model, namely, using residual plots or applying second-order differencing techniques directly to the data [4, 5]. We briefly discuss these methods below. More details can be found in [4, 5].

#### 3.2.1. Residual Plots

Calculating residual plots *after performing* an inverse problem is a common method to determine the appropriate statistical error model. Given the assumed statistical error model (2), for  $j = 1, \dots, N$ , residuals/modified residuals are defined by

$$r_j = \begin{cases} y_j - f(t_j; \hat{\boldsymbol{\theta}}), & \gamma = 0 \\ \frac{y_j - f(t_j; \hat{\boldsymbol{\theta}})}{f(t_j; \hat{\boldsymbol{\theta}})^\gamma}, & \gamma \neq 0. \end{cases}$$

Note that  $r_j$  is an approximate realization of  $\mathcal{E}_j$ . Under the assumption of independence, one can conclude that the residual plot of  $r_j$  vs.  $t_j$  will appear random. If  $\mathcal{E}_j$  have constant variance, the residual plot  $r_j$  vs.  $f(t_j; \hat{\boldsymbol{\theta}})$  will be random. Residual plots with distinct trends such as increasing or decreasing behavior, or fan-like shape indicates the assumptions of constant variance and/or independence are incorrect. One should then re-evaluate the statistical error model selection.

Several sample residual plots are given below in Figs 1-3 for different com-

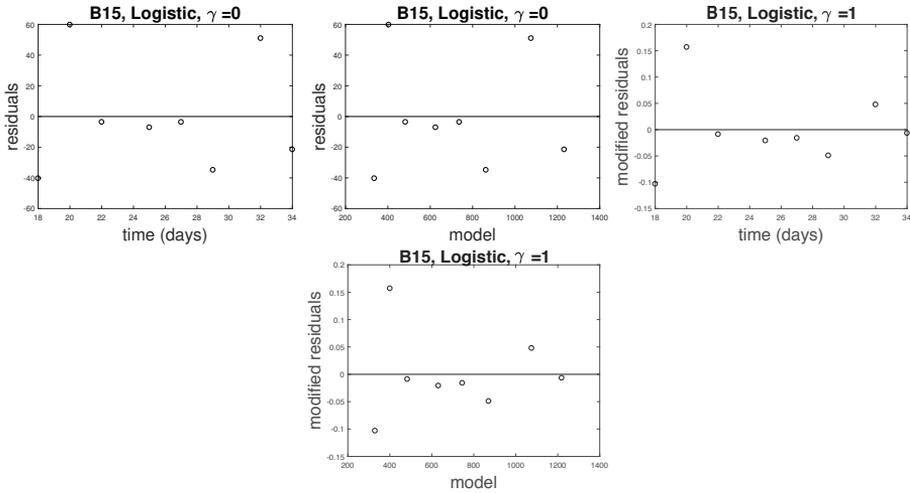


Figure 1: Residuals and modified residuals for breast tumor B15 for  $\gamma = 0$  & 1 using the Logistic model

binations of tumors, statistical error models, and mathematical models. Due to the small sample size in the breast, lung, and HPV tumor data sets, it is difficult to claim one residual plot is more random than another, as seen in the residual plots for the breast tumor logistic model for  $\gamma = 0, 1$  in Fig. 1. In some cases there is an obvious effect when incorrect assumptions in regards to  $\mathcal{E}_j$  are made, such as for the lung tumor fitted by the exponential linear model when  $\gamma = 1$  residual plot in Fig. 2. Moreover, this trend is found in all of the lung tumors among all of the mathematical models for the statistical error model when  $\gamma = 1$ . In general, though, there is rarely a consistent pattern for the selection of a specific statistical error model based on the residual plots for each tumor type and mathematical model, as seen in Fig. 3 for the power law model fit to an HPV tumor data set for  $\gamma = 0, 0.25, 0.5, 0.75, 0.84, 1$ . Thus the selection of  $\gamma$  based on the residual plots is inconclusive due to sparsity of data.

1. Sample residual plots for a selected breast tumors
2. Sample residual plots for a selected lung tumors
3. Sample residual plots for a selected HPV tumors

### 3.2.2. Second-Order Differencing Techniques

To produce residual plots, one has to solve an inverse problem for each  $\gamma$  value. This could be computationally expensive. In addition, if the correct mathe-

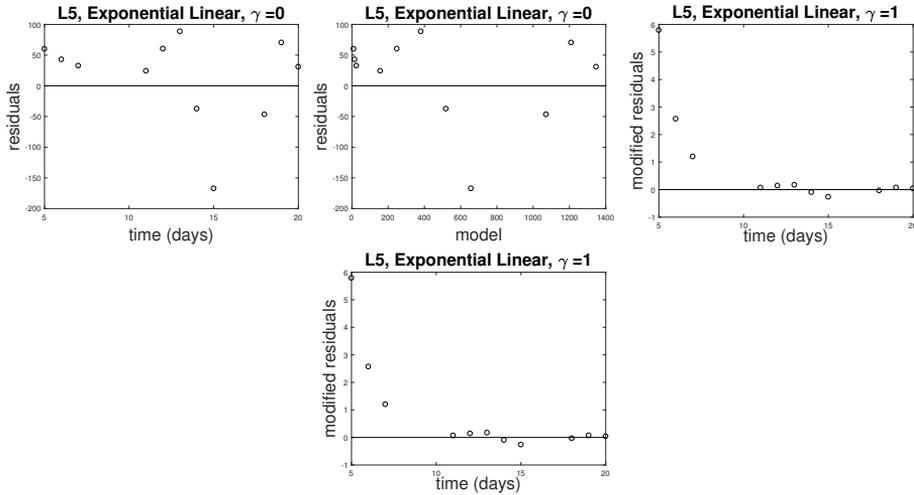


Figure 2: Residuals and modified residuals for lung tumor L5 for  $\gamma = 0$  & 1 using the Exponential Linear model.

mathematical model is not used, residual plots could give inaccurate information, as these plots also depend on the solution of the mathematical model as well as on the chosen  $\gamma$  value. A method that relies *only on the data itself* for identifying the correct observational statistical error model is a *second-order differencing technique* and is described in detail in [4]. This method is found to be more accurate and efficient than using residual plots [4] as well as *not* requiring *prior* solution of inverse problems.

We attempted to use this second order differencing technique directly with several of our larger data sets, namely the HPV tumors. However, due to small sample size, our data sets again failed to yield useful information as to the appropriate statistical models to employ. Thus, even our larger data sets were too small to produce conclusive results for the correctness of statistical models.

### 3.2.3. Evaluation Criteria

As neither of our proposed methods of choosing a statistical error model *a priori* yielded results for our small longitudinal data sets, we examine the effects of the choice of the statistical error model ( $\gamma$  values) on the performance of each mathematical model using four criteria: visual fit, standard errors (SEs), mean square errors (MSEs), and consistency of parameter estimates across  $\gamma$  values. When comparing visual fits, we look for discrepancies between the model fit

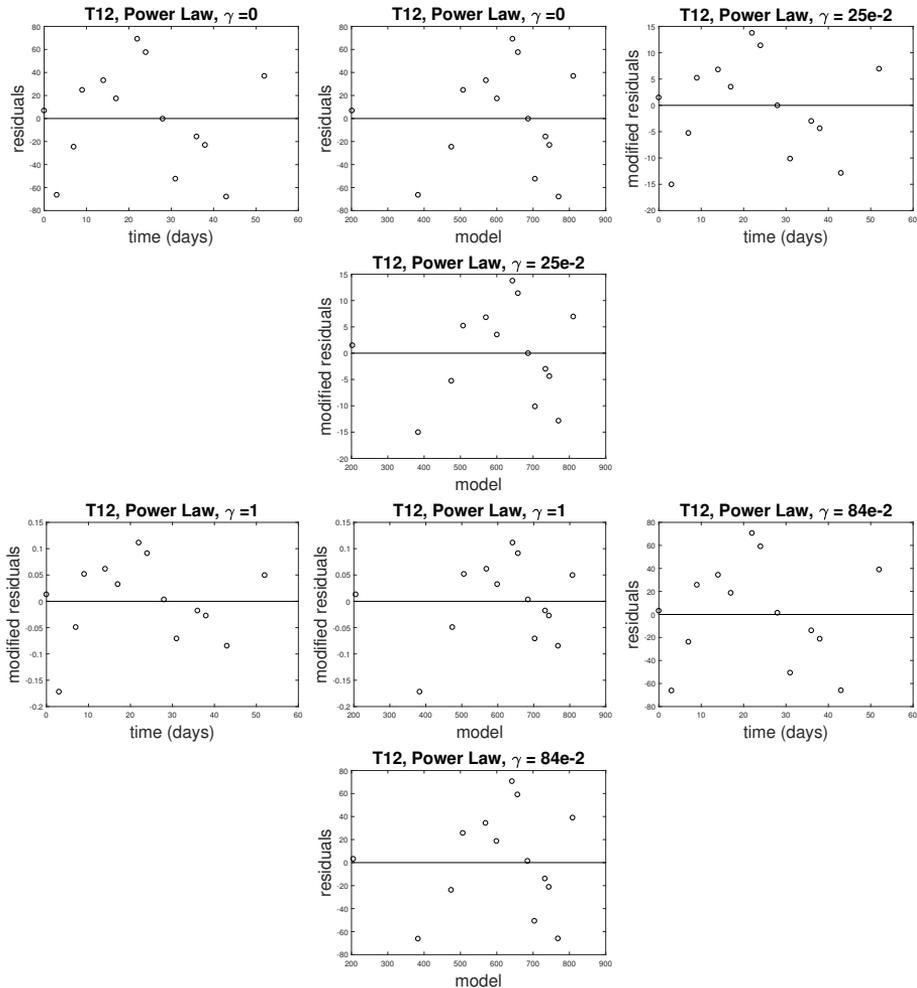


Figure 3: Residuals and modified residuals for HPV tumor T12 for  $\gamma = 0, 0.25, 0.84$  & 1 using the Power Law model.

and the data graphically. If there is little to no discrepancy, we conclude that it is a reasonable visual fit for the data. Standard errors of parameter estimates describe the uncertainty of the estimate. If the SEs of a particular parameter are on the same order of magnitude as the parameter estimate, there is a great deal of uncertainty in that parameter estimate. If the SEs are larger than the parameter estimate, the level of uncertainty in the parameter estimation is too high to draw reasonable conclusions. Mean squared error is a measure of how well the model fits the data. If the MSEs are consistent across statistical models

and SEs for certain parameters are relatively large, we conclude that the model fits are not sensitive to those parameters. If MSEs vary across statistical models, we conclude that certain statistical models capture the dynamics of the data set more accurately than others. We examine the consistency of parameter estimates among the different statistical models to see if changing  $\gamma$  has an effect on the parameter values.

### 3.3. Methods for Selection of a Mathematical Model

Once we have chosen an appropriate statistical error model, we compare mathematical models using a model selection criterion, namely, *the Akaike information criterion AIC*. Assuming that the true mathematical model is known and the measurement errors  $\mathcal{E}_j$ ,  $j = 1, \dots, N$  are independent and identically distributed with mean zero and constant variance  $\sigma^2$  (see Section 3), the AIC for univariate observations is given by

$$\text{AIC} = -2 \ln \mathcal{L}(\hat{\boldsymbol{\theta}}|\mathbf{y}) + 2\kappa_{\boldsymbol{\theta}},$$

where  $\hat{\boldsymbol{\theta}}$  is the estimate of  $\boldsymbol{\theta}$  using the appropriate statistical error model. Because our data set has a small sample size, we will use *the small sample Akaike information criterion AIC<sub>c</sub>*, given by

$$\text{AIC}_c = \text{AIC} + \frac{2\kappa_{\boldsymbol{\theta}}(\kappa_{\boldsymbol{\theta}} + 1)}{N - \kappa_{\boldsymbol{\theta}} - 1}.$$

For the given statistical error model, the mathematical model with the smallest AIC<sub>c</sub> is selected to be the best fit model for the data set. For a detailed discussion on the subject, we refer the reader to [5, 6, 7, 10] and the references therein.

## 4. Results

We fit seven mathematical models to three types of tumor data sets (breast, lung, and skin (HPV) tumors), and used  $\gamma$  values ranging from 0 to 1 for the statistical error model (2). Specifically, we take  $\gamma = 0, 0.25, 0.5, 0.75, 0.84, 1$ . When  $\gamma = 0.84$ , we use the statistical error model as in [9]. This error model was chosen because, in practice, there is a threshold below which accurate measurements are not possible due to detectability limits. Benzekry et al. [9], experimented with different values and concluded that a threshold of  $V = 83\text{mm}^3$  paired with  $\gamma = 0.84$  gave an accurate description of error in the breast

and lung tumor data. The HPV data set did not have a volume measurement below the threshold, thus the statistical error model given by (2) was used when  $\gamma = 0.84$  for the HPV data.

In keeping with conventions in [9], we first fixed  $V_0$  for the lung tumor data; however, we subsequently chose not to fix  $V_0$  for the breast tumor data as we found that estimating this parameter gave a more reasonable visual fit. The lung tumor data was plotted starting at day 0, the day of injection, with the initial condition  $V_0 = 1\text{mm}^3$ . The breast data set was plotted starting at the day of first measurement, using the first measurement volume as the initial estimate. Originally we plotted the breast data starting at day 0, but because of the sparsity of the data, the linear dynamics, and relatively long time interval between day 0 and the first measurement day, we found that plotting using the first measurement day gave a more reasonable visual model fit. In contrast, the lung tumor data fit well visually using day 0 with the fixed initial condition because the first measurement day was much closer (in comparison to the breast data) to day 0.

In the exponential linear model for the lung data set, we provide parameter estimates for  $a_0$ . However, we do not include a parameter estimate for  $a_0$  for the breast or the skin data sets. This is because in both the breast and skin data sets, we start with a relatively large estimate of  $V_0$ , rather than fixing  $V_0 = 1\text{mm}^3$ . When  $V_0$  is large,  $\tau$  is either negative or smaller than the first measurement day, forcing the model to be exclusively linear (Section 2.1.7). Thus it is unnecessary to include the exponential growth rate parameter  $a_0$  as it is not used in our model estimation. With a much larger  $V_0$ , it is reasonable to have exclusively linear growth, as the model implies that exponential growth occurs when the tumor is small (namely, below the  $\tau$  threshold).

## 4.1. Visual Inspection of the Model Fits

We summarize our findings for the various types of tumors based solely on visual fits to data.

### 4.1.1. Breast Tumor Data Sets

For the breast data, the Von Bertalanffy and power law model fits are generally reasonable, but we observe a small change in model fit as  $\gamma$  increases. In Von Bertalanffy, a flattening of the fit curve through the data points in B31 occurs when  $\gamma \geq 0.5$  is used for the statistical error model. For the power law model, the model fit is concave up at  $\gamma = 0$ , flattens out as  $\gamma$  increases in three of

the breast tumors, and flips concavity at  $\gamma = 0.84$  in B31. For B33, the model fit is concave down and the opposite behavior occurs in that as  $\gamma$  decreases the fit flattens out. When using the dynamic carrying capacity model, we see that the fits are overly sensitive to the second data point for most  $\gamma$  values in each of the four breast tumors and for B33 when  $\gamma \geq 0.84$ . As  $\gamma$  increases, the weights on the smaller measurements also increase, which causes an overfit to the cluster of smaller measurements. The logistic, Gompertz, generalized logistic, and exponential linear models give consistently reasonable visual fits regardless of the choice of the statistical error model. In Figure 4, we display plots of the model fits for selected tumor data and  $\gamma$  values.

#### 4.1.2. Lung Tumor Data Sets

When  $\gamma = 0.75$  and  $\gamma = 0.84$ , the logistic model seems to reach a carrying capacity prematurely; this is true for L2, L4, and L9. For the exponential linear model, when  $\gamma \geq 0.75$ , the model fit becomes linear too soon, with a growth rate that underestimates the data in all four lung tumors. When  $\gamma = 1$ , the Gompertz and generalized logistic fit plots do not accurately represent the data for L4, L5 and L9. In the case of the Gompertz plots, the model fit underestimates the volume. For the generalized logistic, the model overestimates the volume. The dynamic carrying capacity, Von Bertalanffy, and power law models have consistent visual fits for all of the statistical models, respectively.

For certain values of  $\gamma$ , the cost function in the IWRLS scheme does not converge to one minimum value, but rather oscillates between two local minima. This is an example of non-uniqueness of solutions in inverse problems. To address this, we choose the parameter set that gives the smallest value of the cost function, and in doing this, we get a more accurate visual fit of the data. Non-convergence of parameter estimates in our minimizer was an issue for all four lung tumors when  $\gamma = 1$  for the logistic model, when  $\gamma = 1$  for the exponential linear model in L4, L5, and L9 and also  $\gamma = 0.75$  for the logistic model (L5). In Figure 5, we display plots of the model fits for selected tumor data and  $\gamma$  values.

#### 4.1.3. Skin (HPV) Tumor Data Sets

Regardless of the statistical error model used, logistic, Gompertz, generalized logistic, dynamic carrying capacity, power law, and Von Bertalanffy models produce consistently good visual fits for three of the four tumors in this data set, namely, T4, T12, and T24. The model fits for T9 exhibit large discrepancies

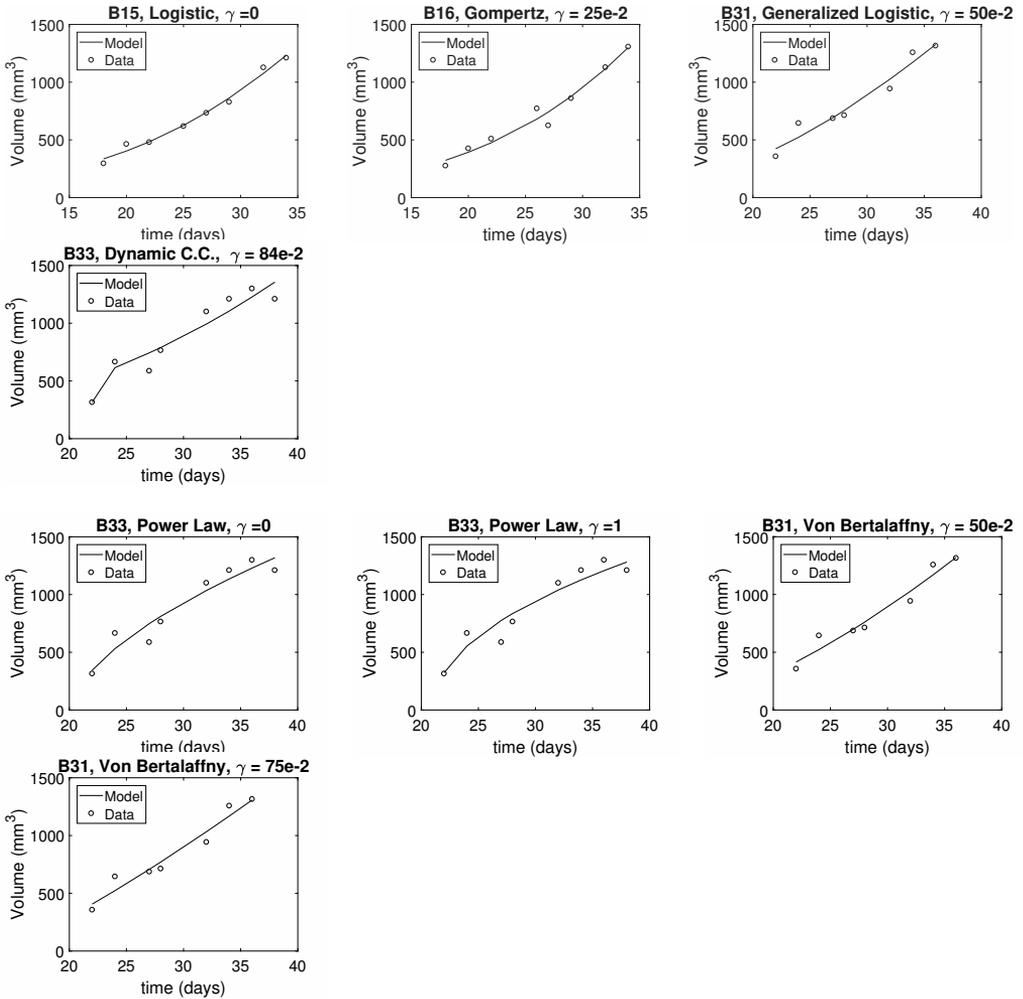


Figure 4: Sample model fits of selected mathematical and statistical models for breast tumor data

between the model fit and the data due to the data's extreme non-monotonic behavior. We note that all mathematical models in this study are monotonic; the dynamics of the data for T9 are fairly sporadic and include an outlier, as seen in the following plots. The exponential linear model does not capture the dynamics of the data for T9 and T12, but gives a better visual model fit for T4 and T24. We assume this is because the latter two data sets exhibit a more linear behavior. In Figure 6, we display plots of the model fits for selected tumor data and  $\gamma$  values.

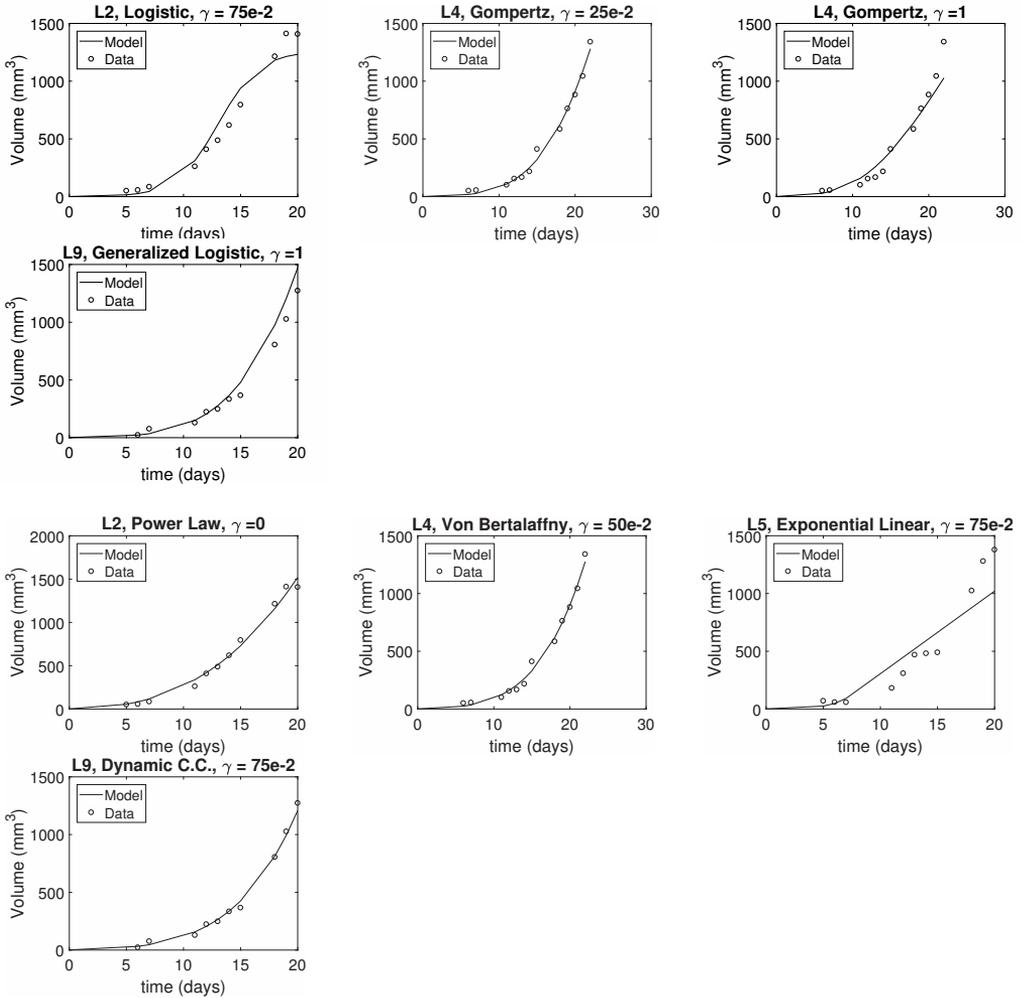


Figure 5: Sample model fits of selected mathematical and statistical models for lung tumor data

### 4.2. Parameter Estimates, Standard Errors and Mean Squared Errors

In order to assess the effect of the statistical error model choice on model fit and certainty in parameter estimation, we examine the change in parameter estimates, standard errors (SEs) and mean squared errors (MSEs) for each of the seven mathematical models and three tumor data sets with respect to the change in statistical error model (values of  $\gamma$ ). For the SEs, we calculate what

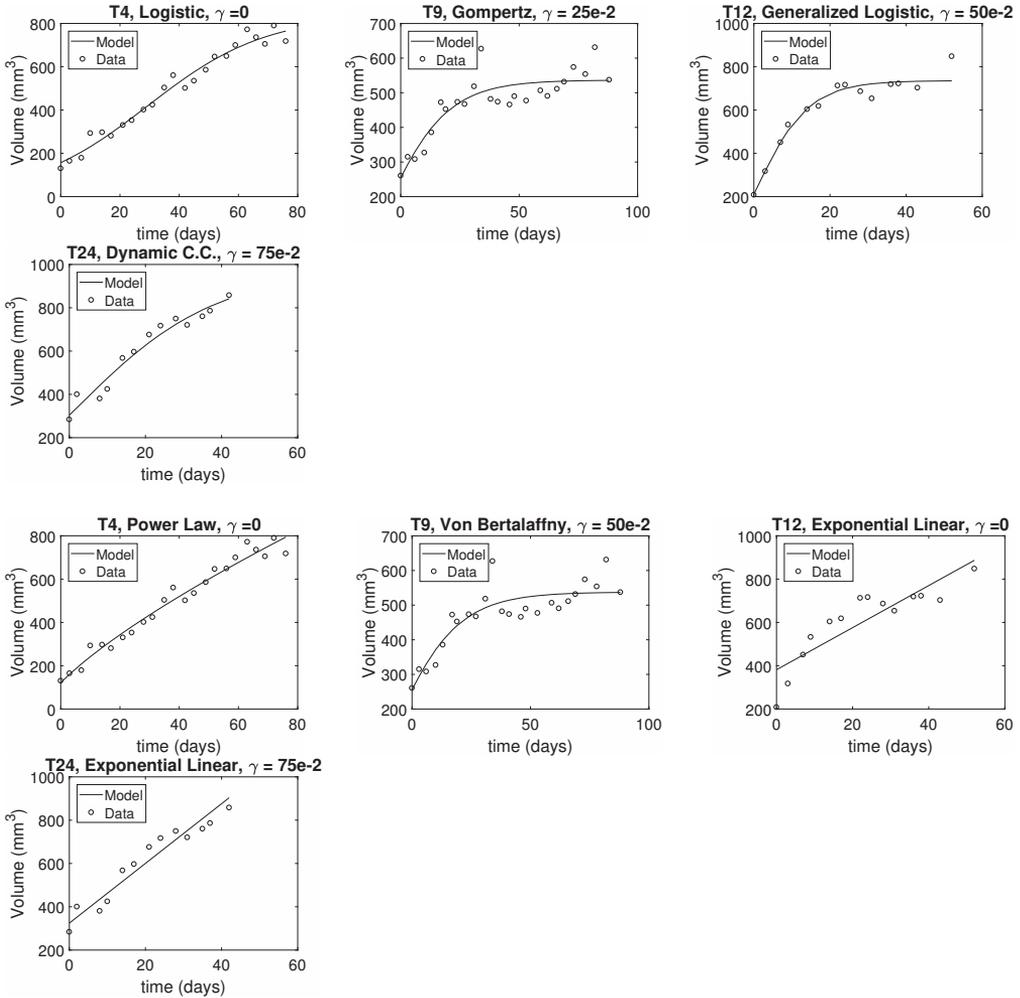


Figure 6: Sample model fits of selected mathematical and statistical models for skin (HPV) tumor data

percentage the SE is of the estimate (i.e.,  $(\frac{SE}{estimate}) 100\%$ ) to better quantify the size of the SE relative to the estimate which indicates the level of uncertainty of the parameter estimate. Further, to quantify how much parameter estimates and MSEs vary depending upon  $\gamma$ , we look at the percent difference in deviation of each parameter estimate (or MSE) from the mean of all parameter estimates (or MSEs) over  $\gamma$ . We present in [3] *representative tables* containing parameter estimates, SEs and MSEs for selected tumor data from each data set. These are given in Tables 1 – 7 of [3] for each of the 7 specific models discussed above.

Here we give a brief summary of the performance of each model with the data sets. More details are given in [3].

### 4.3. Results Summary: Effects of Statistical Models on the Mathematical Model Fits

Following the above observations and those in [3], we discuss the overall effects of statistical models on the mathematical model fits, parameter estimates, standard errors, and mean squared errors for the seven mathematical models considered.

#### 4.3.1. Logistic

The choice of statistical error model does not have much effect on the visual model fit for breast and HPV data sets. In the lung data set, we see that the model fit reaches a carrying capacity prematurely for larger  $\gamma$ . For breast and lung tumor data, the estimates for the carrying capacity  $K$  vary as  $\gamma$  increases, but because the standard errors are large for all  $\gamma$  and the MSEs are consistent among all  $\gamma$ , we conclude the data does not contain sufficient information to estimate  $K$ . We note that for lung tumors, when  $\gamma = 1$  is used for the statistical error model, the cost functional oscillates between two local minima. Spratt et al., [23], found that breast tumors in the developing stages of a clinical study were best modeled by the logistic model. Our findings were generally in agreement with the Spratt findings.

#### 4.3.2. Gompertz

The Gompertz model provides a reasonable visual model fit regardless of  $\gamma$  values for each data set. For the lung data, as  $\gamma$  increases, the MSEs increase, but the parameter estimates and SEs were consistent across statistical models. The breast data showed variance in the  $a$  and  $\beta$  parameters, where each of these had standard errors on the same order of magnitude or larger. These estimates suggest that there is not sufficient data to estimate these parameters. MSEs were consistent for the breast data, suggesting that when coupled with large standard errors, the model fit is not sensitive to  $a$  or  $\beta$ . The Gompertz model fares well in all four criterion for the HPV data. Benzekry et al., concluded Gompertz provided the best model fit for both the lung and breast tumors. In the HPV tumor study the authors chose to exclusively use Gompertz due to the models capability to decrease the growth rate as the tumor increases in

size, [14]. We note that the Gompertz model did not rank as best in any of our findings given in the  $AIC_c$  rankings below.

Data	Par. (SE)	$\gamma = 0$	$\gamma = 0.25$	$\gamma = 0.5$
HPV	$V_0$	254 (32.2)	255 (28.1)	256 (24.6)
tumor	$a$	5.35e-2 (0.02)	5.26e-2 (0.018)	5.18e-2 (0.016)
T9	$\beta$	7.15e-2 (1.87e-2)	7.07e-2 (1.76e-2)	6.98e-2 (1.66e-2)
—MSE—		1.86e+3	1.86e+3	1.86e+3

Data	$\gamma = 0.75$	$\gamma = 1$	$\gamma = 0.84$
HPV	257 (21.5)	258 (18.9)	257 (20.5)
tumor	5.09e-2 (0.015)	5.00e-2 (0.013)	5.06e-2 (0.014)
T9	6.98e-2 (1.56e-2)	6.89e-2 (1.48e-2)	6.86e-2 (1.53e-2)
—MSE—	1.864e+3	1.863e+3	

Table 1: Parameter estimates, SEs, and MSEs using the Gompertz model with HPV tumor T9 data (one of the toughest data sets to fit).

### 4.3.3. Generalized Logistic

All three data sets give reasonable visual fits across all  $\gamma$  values when using the generalized logistic model. We found that the  $a$  and  $\nu$  parameters have standard errors of at least one order of magnitude *larger* than the parameter estimate for all three data sets. Additionally, the breast and lung tumor data sets have large SEs for  $K$ . This implies that there is not sufficient information in the data to properly estimate these parameters. Except for the lung tumors, the MSEs are consistent across  $\gamma$  values, which we interpret as lack of sensitivity of the model to those parameters with large standard errors corresponding to each of the data sets.

### 4.3.4. Dynamic Carrying Capacity

In this model, we found that the HPV and lung tumor data sets have reasonable visual fits. However, we observe overfitting in the plots for the breast data set. The parameter estimates and standard errors for  $V_0$  in the HPV data set were consistent and on a lesser order of magnitude than the estimates, respectively. The MSEs for the HPV data set were consistent as well. All other parameters in all other data sets, across all  $\gamma$ , varied largely and showed great uncertainty in terms of SEs. This again suggests that the data does not contain sufficient information to estimate the parameters, and further, that this is not

an appropriate model for these data sets.

**4.3.5. Power Law**

The visual model fits for this model are reasonable when applied to the lung and HPV data sets. For the breast data, as  $\gamma$  increases, we see a flattening of the curvature of the fit. In some cases, the concavity flips after a certain  $\gamma$  value. The parameter estimates for  $a$  and  $\mu$  vary as  $\gamma$  increases for the breast and HPV data sets, and the standard errors for both parameters are often on the same order of magnitude or larger. Because we have consistent mean squared errors across  $\gamma$ , the magnitude of the standard errors for  $a$  and  $\mu$  imply that the model is not sensitive to these parameters. Benzekry et al., found this model to be useful as a secondary model to fit the lung tumor data. Our findings support those of Benzekry et al., for breast (although the estimated values for  $\mu$  varied in sign) as well as lung tumors but were not particularly useful for the HPV data from [14].

Data	Par. (SE)	$\gamma = 0$	$\gamma = 0.25$	$\gamma = 0.5$
Lung	$a$	1.80 (0.248)	1.67 (0.191)	1.54 (0.143)
tumor L2	$\mu$	0.640 (2.81e-2)	0.657 (2.41e-2)	0.673 (2.02e-2)
—MSE—		3.15e+3	3.28e+3	3.75e+3

Data	$\gamma = 0.75$	$\gamma = 1$	$\gamma = 0.84$
Lung	1.45 (0.106)	1.42 (8.70e-2)	1.41 (0.101)
tumor L2	0.687 (1.67e-2)	0.692 (1.51e-2)	0.694 (1.65e-2)
—MSE—	4.51e+3	5.06e+3	5.12e+3

Table 2: Parameter estimates, SEs, and MSEs using the power law model with lung tumor data.

**4.3.6. Von Bertalanffy**

The visual model fits for the lung and HPV data are consistent and reasonable for all  $\gamma$ . The breast data plots show a flattening of the fit curve through the data points as  $\gamma$  increases. The  $a$  and  $b$  parameter estimates vary greatly across  $\gamma$  values for the breast data. The standard errors for these parameters are often a couple orders of magnitude larger than the estimates for all data sets investigated here. The standard errors for  $\mu$  are larger in magnitude than the parameter estimates for the breast and HPV data sets as well. Aside from lung

data when  $\gamma = 1$ , the MSEs are fairly consistent suggesting that the models are not sensitive to these parameters.

#### 4.3.7. Exponential Linear

The exponential linear model provides a reasonable visual model fit for the breast tumor data. In the HPV data set, we found that this model is not appropriate for two of the tumors, as the data does not follow a linear behavior. Note that the breast and HPV data sets have exclusively linear plots because of the formulation of the model (see 2.1.7 and the note before Section 4.1). For the lung data set, we see both exponential and linear sections of growth. In the lung tumor plots, when  $\gamma \geq 0.75$  the model fit underestimates the volume of the tumor, as it shifts to a linear growth rate too soon. We found that across all  $\gamma$  values, all of the parameter estimates, standard errors, and mean squared errors are consistent and reasonable, except for  $\gamma = 1$  in the lung data set. In this case, the standard error for  $a_1$  was larger than the parameter estimate; we note that non-convergence of the minimizer was an issue here.

Data	Par. (SE)	$\gamma = 0$	$\gamma = 0.25$	$\gamma = 0.5$
Breast data B31 —MSE—	$V_0$	391 (59.1)	395 (54.1)	397 (49.8)
	$a_1$	64.9 (6.96)	64.3 (7.02)	64.0 (7.23)
		5.61e+3	5.62e+3	5.63e+3

Data	$\gamma = 0.75$	$\gamma = 1$	$\gamma = 0.84$
Breast	398 (46.2)	397 (43.1)	398 (45.0)
data B31	63.9 (7.61)	64.1 (8.18)	63.9 (7.79)
—MSE—	5.63e+3	5.63e+3	5.63e+3

Table 3: Parameter estimates, SEs, and MSEs using the exponential linear model with breast tumor data.

#### 4.4. Model Comparison

Based on our observations, the choice of statistical error model does not have a predictable or meaningful effect on model fit, parameter estimates, or SEs for data with a small sample size. We see that almost all of the data sets and mathematical models considered in this study revealed  $\gamma \leq 0.25$  corresponded to a reasonable choice of statistical error model. Thus, we suggest that for tumor growth data with small sample size, using simpler statistical models like

the absolute error model is sufficient. With this assumption, we compared seven mathematical models to select the ones that most accurately fit each of the three tumor data sets. We use a *small sample Akaike information criterion*  $AIC_c$  and let  $\hat{\theta}$  be the ordinary least squares estimate (Subsection 3.3). The results of that comparison are listed below.

Mathematical model	B15	B16	B31	B33
Logistic	<b>68.6775</b>	76.9874	74.1706	<b>83.8831</b>
Gompertz	68.7108	76.926	73.5544	84.3769
Generalized logistic	78.0108	86.3675	87.5746	93.216
Dynamic carrying capacity	76.6403	86.22949	85.2901	93.7107
Power law	68.70729	<b>76.91341</b>	<b>73.5232</b>	85.0113
Von Bertalanffy	78.0428	86.2467	87.5233	93.7331
Exponential linear	75.0403	81.24118	74.4261	86.1734

Table 4:  $AIC_c$  scores for four breast tumor data using absolute error statistical model

Mathematical model	L2	L4	L5	L9
Logistic	89.9735	105.5746	101.1007	84.9384
Gompertz	87.1341	93.8046	92.3032	71.8832
Generalized logistic	<b>85.6942</b>	97.5030	96.2457	76.2243
Dynamic carrying capacity	91.0634	95.7695	95.6755	72.4129
Power law	94.0932	<b>92.4395</b>	<b>91.8300</b>	<b>69.4471</b>
Von Bertalanffy	91.4186	96.1062	95.7513	73.7327
Exponential linear	87.3220	106.8956	99.3937	87.6840

Table 5:  $AIC_c$  scores for four lung tumor data using absolute error statistical model

Mathematical model	T4	T9	T12	T24
Logistic	<b>167.5015</b>	<b>187.84</b>	112.5700	<b>101.5624</b>
Gompertz	167.6620	187.899	111.1681	102.24
Generalized logistic	170.389	190.749	115.21	105.5
Dynamic carrying capacity	170.621	189.065	<b>109.10</b>	106.57
Power law	170.457	195.415	113.02	104.2237
Von Bertalanffy	170.6813	190.814	113.82	106.59
Exponential linear	174.701	201.077	131.65	108.31

Table 6:  $AIC_c$  scores for four HPV tumor data using absolute error statistical model

Based on the  $AIC_c$  scores given in Tables 4-6, the logistic, power law, and Gompertz models give the better fits for the breast tumor data. For the lung tumor data, the power law, Gompertz, and dynamic carrying capacity are the top three mathematical models that provide the better fits to the data. And finally, for the HPV tumor data, in general the logistic and Gompertz models give the better fits to the data.

## 5. Discussion and Conclusions

We fit seven tumor growth mathematical models (logistic, Gompertz, generalized logistic, dynamic carrying capacity, power law, Von Bertalanffy and exponential linear) to three types (breast, lung and skin) of tumor data sets. We thoroughly examined the performance of the mathematical models and the effects of the choice of the statistical models ( $\gamma$  values) using four criteria. The  $\gamma$  values we used range from 0 to 1. To determine the appropriate statistical error models, we analyzed the visual model fits, the consistency of parameter estimates, standard errors (SEs) and mean squared errors (MSEs).

In our study, we discovered that if a data set is small, simple residual plots cannot effectively provide information to determine if a statistical error model is appropriate. Neither can second order differencing-based techniques be used to choose the appropriate statistical error model. Both of these methods require the ability to see a pattern or to determine there is no pattern. The small sample size makes any assertions on patterns or randomness unreliable at best since the addition or removal of a single data point can often make one come to an opposite conclusion than originally determined. Therefore, we have to examine the impact of the choice of statistical error model on the overall model

fit (parameter estimates, SEs, MSEs,  $AIC_c$ ) instead to determine the best choice for statistical error model.

In general, we observed that when  $\gamma$  is large (often  $\gamma \geq 0.75$ ) visual model fits begin to look inaccurate; similarly, SEs and MSEs tend to increase to unreasonable sizes. Therefore, for small sample sizes, a simpler statistical error model like absolute error is sufficient as the model fits either deteriorated or did not improve when using other statistical models ( $\gamma > 0$ ) due to the sparsity in the data.

Assuming that absolute error is the appropriate statistical error model, we used likelihood based model selection criterion ( $AIC_c$ ) to determine the mathematical models that best fit each of the three types of tumor data. According to our findings, logistic, power law and Gompertz models produce the best fit for the breast tumor data; power law, Gompertz and dynamic carrying capacity are suited for the lung tumor data; and logistic and Gompertz models provide the best fits for the skin (HPV) tumor data. Based on our proposed evaluation criteria, we found that the Gompertz model fit the HPV data best (despite not being featured as a selection based on  $AIC_c$ ), the power law model fit the lung tumor data best, and there was not a best model that consistently fit the breast tumor data set based on our criteria due to the small sample size.

In general, to estimate mathematical model parameters, the amount of data and information content in the data are crucial. Using the breast and lung tumor data sets, we observed that one can only estimate at most two parameters with a reasonable accuracy. In the case of skin (HPV) tumor data sets, one can estimate up to three parameters due to the relatively larger sample size of the data. The lack of information in the data increases the uncertainty in parameter estimation. For example, we observed for models that have many parameters, SEs were very large. In some cases, for models like the logistic model, the parameter estimates for carrying capacity  $K$  were also very inconsistent and the standard errors were very large. Even though some parameters were fixed (for example initial condition  $V_0$ , in lung tumor data), there were no significant improvements. Therefore, the data was not collected for a sufficiently long period for the tumors to reach the carrying capacity and our estimates of carrying capacity were uncertain and inconsistent as a result. In addition, it was only the mathematical models that were not sensitive to their parameters for the time intervals considered that provided consistently good visual fits and MSEs regardless of large SEs for some parameters. Sensitivity analysis results (not included here) also confirm this, as most of the sensitivity magnitudes were close to zero. Models like exponential linear generally perform better for these short time interval data sets as only the linear part is fitted and the data

contains the information to estimate the few parameters in the linear model.

Based on the above observations, we believe future research is necessary to determine the minimum amount of data needed to accurately estimate parameters in a mathematical model and for the statistical error model to affect the model fits. Specifically, further investigation is desirable on the experimental design in regards to measurement frequency and time span of the tumor measurements. In addition, more research is necessary to explore the minimum number of observations required for methods like residual plots to provide useful information regarding the appropriate statistical error model and for the second order differencing based technique to be valid. Both of the articles we used to gather the data sets were limited by their time scales and frequency of measurements, which limited the accuracy of model parameter estimation [9, 14].

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